Detrimental Effects of Neem Seed on Different Life Stages of Red Flour Beetle, *Tribolium castaneum*

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Abstract: Fine Neem Seed Powder (NSP) was tested in three dozes, viz. 0.5%, 1.0% and 2.0% (w/w) for determination of its toxicity and detrimental effects on life stages of red flour beetle, *Tribolium castaneum*. Fifty gram treated and untreated (control) wheat was kept in glass jars and 10 pairs of the beetles were released in each. Mortality count was made after 72 hours. LD_{50} calculated as 1.841% NSP in wheat (w/w). NSP served as an Insect Growth Regulator (IGR). It was observed that at 0.5% dose, the number of larvae was not very different to control but the weight of larvae was lesser. At 1.0% dose, the number and weight of larvae were significantly reduced. At 2.0% dose, both the number and weight of larvae for served pre-and adults were reduced remarkably. The insect growth inhibition was increased by increasing the dose of NSP.

Keywords: Neem seed powder, antifeedant activity, red flour beetle.

INTRODUCTION

Grains become infested by insect pests during storage rendering them unfit for human consumption. Red flour beetle, Tribolium castaneum (Herbst), is the most abundant and injurious pest of stored grains. It is one of the major pests of wheat in warehouses and flour mills at warm and tropical regions of the world, as reported by various researchers including Semple [1], Arnaud et al. [2], Zettler [3] and Trematerra & Sciarretta [4]. Several methods are being used to overcome the problem. Kostyukovsky [5] emphasized on the need for searching safe and natural management methods as alternative to pesticides, considering the increasing concern about health and environment. Utilization of natural plant products, e.g., mixing of neem materials to save grains from insect pests is an old-age practice in some parts of the world. Natural products of plants have been tested by various researchers for their insecticidal properties against insect pests, such as, Dennis [6], Hossafay & Faisal [7], Emeasor et al. [8], Tooba et al. [9], Iqbal et al. [10], Sarwar & Sattar [11] and Sarwar et al. [12]. There is still need to search easily available bioactive plant materials for grain protection from insect pests during storage. All parts of neem, Azadirachta indica, have some biologically active components that exert detrimental effects on insect population. Neem derivatives are generally not hazardous to agro-ecosystem, as well as insect resistance is not developed like synthetic insecticides.

Various researchers have worked on different aspects of *Azadirachta indica*. They include Meisner & Ascher [13], Naqvi *et al.* [14, 15], Jilani *et al.* [16], Schmutterer [17], Sharma [18], Xie *et al.* [19], Akhter *et al.* [20], Mordue *et al.* [21], Isman [22], Khan [23, 24], Ahmad *et al.* [25], Khan & Ahmad [26], Koul [27], Athanassiou *et al.* [28] and Iqbal [29]. However, there is a need for determination of more bio active plants for management of red flour beetle as identified by Wong [30]. This research was conducted to evaluate detrimental effects of neem seed powder (NSP) against red flour beetle, *Tribolium castaneum* (Herbst). The research may enrich the existing knowledge on effective, safe and environmental friendly protection of stored grains by the indigenous plants.

MATERIALS AND METHODS

Rearing Technique

Insects were reared in laboratory at $30^{\circ}C \pm 1^{\circ}C$ temperature with relative humidity of $60\% \pm 5\%$ on whole wheat grains as food medium in glass jars, covered with muslin cloth. Ten-day old adults were used for all the trials.

Test Neem Material

Fine neem seed powder (NSP) was used in all the trials. Neem seeds were collected and dried under shade. Dried seeds were ground in an electrical grinder to a fine powder of 30 meshes. This fine NSP was tested in three doses of 0.5%, 1.0% and 2.0% (w/w).

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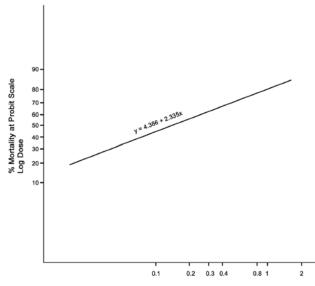


Figure 1: Probit Mortality curve of T castaneum.

Treatment Method

Fifty gram of treated and untreated (control) wheat was kept in 450-gm capacity jars. Twenty, 10-day old adults of red flour beetle, *Tribolium castaneum*, were released in each jar. Insects were removed from the jars after 72 hours and mortality was noted. All the trials were replicated five times. Observations were regularly recorded to check the growth of the insects. Total count and weight were measured after 15 days for larvae, 25 days for pupae and 40 days for adults. All

the tests were performed under the same temperature and humidity. All the data were analyzed statistically.

RESULTS AND DISCUSSION

Fine NSP was tested in three doses of 0.5%, 1.0% and 2.0% (w/w) against adult red flour beetles. Mortality count was made after 24 hours of treatment. Adult mortality was calculated with Probit Analysis after Varely et al. [31]. Probit line is shown in Figure 1. Probit regression equation calculated was Y = 4.526685 +2.335113 (X - (6.150449E - 02)) whereas heterogeneity factor (Chi Square) was 8.330109 (Table 1). Probit and Lethal Dose (LD) values are presented in Table 1. The log LD₅₀ at 72 hours treatment was found to be 1.84%; whereas, potential mortality was higher in 1% treated wheat. It could be probably due to the fact that higher dose because of high antifeedant effect prevented the organism to consume the toxic material. Contrarily, the moderate doses were consumed more and therefore caused higher mortality effect over the generation (Table 3). Survival and effect of NSP on larval and pupal weights are shown in Tables 2 & 3. It was found that NSP exerted weight reduction effect on larval, pupal and adult stages as average larval, pupal and adult weights were found to be 0.386 gm, 0.296 gm and 0.190 gm in control; 0.272 gm, 0.142 gm and 0.090 gm in 0.5% treated; 0.190 gm, 0.044 gm and 0.023 gm in 1.0% treated; and 0.112 gm, 0.022 gm and 0.002 gm in 2% treated, respectively. These results

Table 1: Toxicity of NSP Against Tribolium castaneum (Adults) after 72 Hours

Compound	Conc. (%)	Dose (gm)	Mortality (%)	SD	SE	Mean ± SE x Sig. value at 95% confidence limit				
Control	0.00	0.00	02.00	2.70	1.22	2±2.4				
NSP	0.50	0.25	12.00	5.70	2.55	12±5.0				
NSP	1.00	0.50	24.00	8.22	3.67	24±7.2				
NSP	2.00	1.00	56.00	9.62	4.30	56±8.4				

Regression equation: Y = 4.526685 + 2.335113 (X – (6.150449E – 02)). Heterogeneity factor: Chi Square χ^2 = 8.330109.

	Lethal Dose	Standard Error	Fudicial Limits (95%)				
LD 10	0.521	0.037	0.443	0.613			
LD 30	1.102	0.022	1.007	1.208			
LD 50	1.841	0.025	1.632	2.077			
LD 70	3.074	0.410	2.543	3.717			
LD 90	6.504	0.068	4.781	8.849			
LD 99	18.318	0.107	11.349	29.565			

Conc.	Poplication	Lai	vae	Pup	bae	Adults			
(w/w)	Replication	Count	Wt. (gm)	Count	Wt. (gm)	Count	Wt. (gm)		
	1 st	150.00	0.39	112.00	0.24	98.00	0.141		
Control	2 nd	128.00	0.21	105.00	0.22	102.00	0.181		
	3 rd	161.00	0.45	98.00	0.19	89.00	0.190		
Control	4 th	165.00	0.37	147.00	0.38	120.00	0.210		
	5 th	210.00	0.51	193.00	0.45	149.00	0.230		
	Total	814.00	1.93	655.00	1.48	558.00	0.952		
	1 st	115.00	0.28	41.00	0.10	31.00	0.065		
	2 nd	142.00	0.30	79.00	0.17	48.00	0.079		
0.5%	3 rd	171.00	0.39	82.00	0.15	59.00	0.093		
	4 th	143.00	0.21	67.00	0.13	58.00	0.103		
	5 th	125.00	0.18	78.00	0.16	62.00	0.110		
	Total	696.00	1.36	347.00	0.71	258.00	0.450		
	1 st	72.00	0.21	48.00	0.06	22.00	0.041		
	2 nd	102.00	0.24	40.00	0.06	16.00	0.010		
1.0%	3 rd	65.00	0.18	14.00	0.02	4.00	0.002		
1.0%	4 th	93.00	0.21	55.00	0.10	26.00	0.043		
	5 th	55.00	0.11	21.00	0.03	13.00	0.018		
	Total	387.00	0.95	178.00	0.27	81.00	0.114		
	1 st	16.00	0.05	8.00	0.01	1.00	0.001		
	2 nd	28.00	0.09	10.00	0.04	3.00	0.004		
2.0%	3 rd	7.00	0.02	5.00	0.01	1.00	0.001		
∠.0%	4 th	25.00	0.09	8.00	0.02	2.00	0.002		
	5 th	31.00	0.12	12.00	0.03	5.00	0.005		
	Total	107.00	0.37	43.00	0.11	12.00	0.013		

Table 2: Effect of Neem Seed Powder on Immature Stages of Tribolium castaneum

show an increase in weight reduction with in NSP concentrations. This phenomenon could be due to the antifeedant activity of neem as indicated by Sharma et al. [32] and Khan [23]. Weight reduction could also be due to growth retardation and difference in metabolism. Growth retardation by neem derivatives has been reported by Sharma [18] and Khan & Ahmed [26]. The present findings on neem against the beetle are in line with the previous reports. As indicated before, NSP exerted acute mortality in adults. Killing potential (k factor) for treated medium reared beetles observed was 0.49 for 0.5%, 0.99 for 1% and 1.8 for 2% as compared to the untreated (control) where k factor was found 0.16. Similar findings on toxicity of neem seed against T. castaneum and T. cofusum are reported by Ahmad et al. [25] and Khan & Ahmad [26]. The present findings on detrimental effects of neem seed powder

may be a beneficial addition in the work (on plant products) conducted by earlier researchers. Some researchers evaluated plants powder for their effectiveness against stored grain pests. Al-Moajel [33] prepared powders from parts of different plant species and found (62%) mortality at 6% concentration against Trogoderma granarium; Silva et al. [34] evaluated up to 99.3% mortality of Sitophilus zeamais by powdered plants; Emeasor et al. [8] found plant seed powders very toxic at 1 and 2 gm/ 20 gm seeds against Callosobruchus maculates. The results of the study indicated that the neem seed has good biological activity against insect and fine NSP may be used for stored grain treatment as dusting and bag treatment as well. The use of neem seed powder (NSP) would not need any standardization; however, Sivakumar et al. [35] advised use of appropriate machinery, e.g.,

Table 3: Killing Factor and Potential Mortality Effects on Various Life Stages of Tribolium castaneum Reared on Neem Seed Treated Media Seed Treated Media

	Control *							C	0.5% **				0.1% ***					0.2% ****								
	NL	\textbf{SI}_{LP}	NP	SI_{PA}	NA	NE	SI_{EL}	NL	\textbf{SI}_{LP}	NP	\mathbf{SI}_{PA}	NA	NE	SI_{EL}	NL	SI_{LP}	NP	SIPA	NA	NE	SI_{EL}	NL	\textbf{SI}_{LP}	NP	\mathbf{SI}_{PA}	NA
Population	814		715		558	814		696		347		258	814		387		178		81	814		107		43		12
Dying Interval		99		159			118		349		89			427		209		97			707		64		31	
Mortality (%)		12.2		19.3			14.5		42.9		10.9			52.5		25.7		1.0			87.0		7.86		3.8	
Successive Mortality (%)		12.2		22.0			14.5		50.0		26.0			52.5		54.0		54.5			87.0		60.0		72.0	
Successive Survival (%)		87.8		78.0			85.5		50.0		74.0			47.5		46.0		45.5			13.0		40.0		28.0	
Fraction Survival		0.88		0.78			0.86		0.50		0.74			0.475		0.460		0.455			0.13		0.40		0.28	
Log Population	2.91		2.85		2.75	2.91		2.84		2.54		2.41	2.91		2.59		2.25		1.91	2.90		2.03		1.63		1.08
K-Value		0.06		0.10			0.06		0.30		0.13			0.31		0.34		0.34			0.87		0.40		0.55	

	Σ Mortality	Σ Mortality (%)	Generation Survival	K-factor
*	258	31.5	0.680	0.16
**	556	68.3	0.320	0.49
***	733	90.11	0.099	0.99
****	802	98.66	2.170	1.82

NE: Number of eggs.

NL: Number of larvae.

NP: Number of pupae.

NA: Number of adults.

SI_{EL}: Standard interval between eggs to larvae.

SI_{LP}: Standard interval between larvae to pupae.

SI_{PA}: Standard interval between pupae to adults.

decorticator, seed/kernel crusher and pulverizer that may be needed for processing plants at village level.

REFERENCES

- Semple RL. Problems relating to pest control and use of pesticides in grain storage: The current situation in ASEAN and future requirements. Pesticides and Humid Tropical Grain Storage System: ACIAR Proc 1986; 45-47.
- [2] Arnaud L, Lognay G, Verscheure M, Leenaers L, Gaspar C, Haubruge E. Is dimethyldecanal a common aggregation pheromone of *Tribolium* flour beetles? J Chem Ecol 2002; 28(3): 523-32. http://dx.doi.org/10.1023/A:1014587927784
- [3] Zettler JL. Pesticide resistance in *Tribolium castaneum* and *T. confusum (Coleoptra: Tenebrionidea)* from flour mills in the United States. J Eco Entomol 1991; 89(3): 763-67.
- [4] Trematerra P, Sciarretta A. Spatial distribution of some beetles infesting a feed mill with spatio-temporal dynamics of *Oryzaephilus surinamensis*, *Tribolium castaneum* and *Tribolium confusum*. J Stored Prod Res 2004; 40(4): 363-77. http://dx.doi.org/10.1016/S0022-474X(03)00027-4
- [5] Kostyukovsky M, Rafaeli A, Gileadi C. Activation of octopaminergic receptors by essential oil constituents isolated from aromatic plants: possible mode of action against insect pests. Pest Management 2002; 58(11): 1101-106. http://dx.doi.org/10.1002/ps.548

- [6] Dennis SH. Pest of stored products and their control. London: Bet Haven Press 1990; pp. 219-220.
- [7] Hossafay ME, Faissal FA. Screening certain products and organic acid for controlling vorrao mites *Vorrao jacobsoni* (Oudemans). Shashpa 1994; 1(1): 56-62.
- [8] Emeasor KC, Ogbuji RO, Emosairue SO. Insecticidal activity of some seed powders against *Callosobruchus maculates* (F) (Coleoptera: Bruchidae) on stored cowpea. Zeitschrift fur Pflanzenkrankheiten und Pflanzenschutz 2005; 112(1): 80-87.
- [9] Tooba H, Usman NF, Abbas T. Screening of plant leaves as grain protectants against *Tribolium castaneum* during storage. Pak J Bot 2005; 37(1): 149-53.
- [10] Iqbal J, Jilani G, Aslam M. Growth inhibiting effects of plant extracts against the grain moth, *Sitotroga cerealella* (Oliv.) (*Gelechiidae: Lepidoptera*). Pak J Zool 2010; 42(5): 597-601.
- [11] Sarwar M, Sattar M. Appraisal of different plant products against *Trogoderma granarium* Everts to protect stored wheat – a laboratory comparison. Nucleus 2012; 49(1): 5.
- [12] Sarwar M, Ashfaq M, Ahmad A, Randhawa MAM. Assessing the potential of assorted plant powders on survival of *caloglyphus* grain mite (*acari: acaridae*) in wheat grain. Int J Agric Sci Bioresour Eng Res 2013; 2(1): 1-6.
- [13] Meisner J, Ascher KRS. Insect growth-regulating (IGR) effects of neem products on *Spodoptera littoralis*. Schriftenr Ges Tech Zusammenarbeit 1984; 161: 345-52.
- [14] Naqvi SNH, Tabassum R, Zia N, Nurulain SM. Toxicity and residual effect of neem extract (factor C) against stored grain pest *Callosobruchus analis*. Pak J Zool 1990; 22: 271-77.

- [15] Naqvi SNH, Ahmed SU, Mohammad FA. Toxicity and IGR effects of two neem products against *Aedes aegypti (PCSIR Strain)*. Pak J Pharm Sci 1991; 4: 71-76.
- [16] Jilani G, Noorullah, Ghiasuddin. Repellency of some plant extracts against *Tribolium castaneum* (Herbst) (*Coleoptera: Tenebrionidae*). Pak Entomol 1991; 13(1-2): 5-8.
- [17] Schmutterer H. Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. A Rev Ent 1990; 35: 271-97. http://dx.doi.org/10.1146/annurev.en.35.010190.001415
- [18] Sharma GK. Growth inhibitor activity of *Azadirachta indica* on *Corcyra cephalonica*. Phytoparasitics 1992; 20: 251-58.
- [19] Xie YS, Field PG, Isman MB. Repellency and toxicity of azadirachtin and neem concentrates to three stored-product beetles. J Eco Entomol 1995; 88(4): 1024-31.
- [20] Akhtar K, Naqvi SNH, Azmi MA. Determination of toxicity, fecundity and emergence effects of neem factor (NFQ) against *Callosobruchus analis* (*L.*). Zeithschritt Fur Angewandte Zoologie 1996; 80(2): 141-54.
- [21] Mordue AJ, Nisbet AJ, Nasiruddin M, Walker E. Differential thresholds of azadirachtin for feeding deterrence and toxicity in locust and aphid. Entomologia Exp et Application 1996; 80: 69-72. http://dx.doi.org/10.1111/j.1570-7458.1996.tb00887.x
- [22] Isman MB. Neem insecticides. Pesticde Outlook 1997; 32-38.
- [23] Khan MF. Determination of resistance in cotton pest *Earias febia* (Stoll) against local neem product as compared to monocratophos and cypermetharin [Ph.D. thesis]. Karachi: University of Karachi 1998.
- [24] Khan MF, Ahmed SM. Toxicity of neem fruit extract and seed oil against *Oxycarenus* bug of cotton crop. Acta Biologica Caracoviensia, Series Zoologia 2000; 42: 17-21.
- [25] Ahmad I, Ali RR, Tabassum R, Azmi MA, Naqvi SNH, Khan MF. Toxicity determination of two pyrethroids, Karate (cyhalothrin) and delatamethrin as compared to neem extract on *Tribolium castaneum*. J Exp Zool India 2001; 4(1): 169-73.
- [26] Khan MF, Ahmed SM. Comparative toxicity of neem fruit extract and cypermethrin against *T.cofusum Hbbst* and *Papilio demoleeus Linn*. Philippine J Sci 2003; 132: 109-14.

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[27] Koul O. Biological activity of volatile di-n-propyl disulfide from seeds of neem, Azadirachta indica (Meliaceae), to two species of stored grain pests, Sitophilus oryzae (L.) and Tribolium castaneum (Herbst). J Eco Entomol 2004; 97(3): 1142-47. http://dx.doi.org/10.1603/0022-

0493(2004)097[1142:BAOVDD]2.0.CO;2

- [28] Athanassiou CG, Kontodimas DC, Kavallieratos NG, Veroniki MA. Insecticidal effect of *Neemazal* against three storedproduct beetle species on rye and oats. J Eco Entomol 2005: 98(5): 1733-38. <u>http://dx.doi.org/10.1603/0022-0493-98.5.1733</u>
- [29] Iqbal J. Growth inhibiting and deterrent effects of plant extracts on major insect pests of stored grains [PhD thesis]. Rawalpindi: University of Arid Agriculture 2006.
- [30] Wong KKY, Signal FA, Campion SH, Motion RL. Citronella as an insect repellent in food packaging. J Agri Food Chem 2005; 53(11): 4633-36. <u>http://dx.doi.org/10.1021/ijf050096m</u>
- [31] Varely GC, Gradwell GR, Hassell MP. Insect population ecology: An analytical approach. Blaclell Sci Pub 1975; 6-8.
- [32] Sharma HC, Leuscher K, Sankaram AVB, Gunaserhar D, Marthandamurthi M, Sultan N. Insect antifeedants and growth inhibitors from *Azradirachta indica*. Proc 2nd Int Neem Conf 1983. 1984; pp. 291-320.
- [33] Al-Moajel NH. Testing some various botanical powders for protection of wheat grain against *Trogoderma granarium* Evert. J Biol Sci 2004; 4(5): 592-97. <u>http://dx.doi.org/10.3923/jbs.2004.592.597</u>
- [34] Silva G, Orrego O, Hepp R, Tapia M. Search for plants with insecticidal properties for *Sitophilus zeamais* control in stored corn. Pesquisa-Agropecuaria-Brasileir 2005; 40(1): 11-17. http://dx.doi.org/10.1590/S0100-204X2005000100002
- [35] Sivakumar SS, Palanisamy PT, Vardharaju N, Gothandapani L, Swaminathan KR. Machineries for neem processing. In: Singh RP, Chari MS, Raheja AK, Kraus W, Eds. Neem and Environment, Proc World Neem Conf, Bangalore. New Delhi: Oxford & IBH Publ 1991; Vol. 2.