# Parasite Inspection in Five Commercially Important Oyster Species (*Mollusca: Bivalvia*) of Pakistan

Nuzhat Afsar<sup>1,\*</sup>, Ghazala Siddiqui<sup>2</sup> and David Roberts<sup>3</sup>

<sup>1</sup>Institute of Marine Science, University of Karachi, Karachi-75270, Pakistan <sup>2</sup>Center of Excellence in Marine Biology, University of Karachi, Karachi 75270, Pakistan <sup>3</sup>School of Biological Sciences, Queen's University, Belfast, UK

**Abstract**: A total of five native oyster species namely; *Crassostrea gryphoides, C. madrasensis, C. glomerata, Ostrea nomades* and *Saccostrea cucullata* were collected from three sites: Buleji of Sindh coast; Hub and Jiwani of Balochistan coast, Pakistan. Parasitic inclusion was present only in histological sections of *O. nomades* and *S. cucullata* at Buleji. Individuals of *O. nomades* found to be infected with protozoan parasite *Nematopsis* species and *Proctoeces* sp. trematode parasite. Additionally, unknown parasitic cyst was observed at outer epithelium in a male gonadal section of *S. cucullata*. There is no previous record in literature of pathogens in oyster species at Buleji.

Keywords: Nematopsis, Northern Arabian Sea, Ostrea, Saccostrea, Crassostrea, Trematode, parasite.

## INTRODUCTION

Over the past 60 years number of pathogens, related diseases and mortalities have been reported in bivalves across the globe [1-7]. The main biological disease-causing agents in marine bivalves are viruses, bacteria, fungi, protists, digenean trematodes, polychaetes and copepods [8]. Parasitic diseases pose a serious threat to the world's oyster aquaculture industry and to oyster fisheries. Historically, some diseases are of regulatory significance [9, 10] have been associated with Ostrea spp. and Crassostrea spp. and account for major commercial losses and limit expansion of oyster culture. Pakistan has a sizeable diversity of native species of oysters [11] which can provide a potential resource for the development of mariculture in Pakistan [5]. However, although they are not exploited commercially, oyster stocks in Pakistan have become depleted and their natural spat fall is very low [12]. Like elsewhere in the world the study of parasites and disease distressing oysters is important due to their potential economic impacts on the management of natural stocks and aquaculture [5-6]. Siddigui et al., [5] reported the Nematopsis spp. in O. nomades and C. belcheri from Keamari Sea wall area adjacent to Manora Channel along the Sindh coast of Pakistan. The aim of the present study was to broaden this previous study to other species and sites as the diseases of regulatory significance not found in native oyster species which make them exclusively important from the mariculture point of view for commercial exploitation.

#### METHODS

Samples were procured, between January 2012 and March 2012 at low tides (Table 1). Five oyster species, Crassostrea gryphoides (Schlotheim, 1813), C. madrasensis (Preston, 1916), C. glomerata (Gould, 1850), Ostrea nomads (Iredale, 1939) and Saccostrea cucullata (Born, 1778) were sampled from the intertidal zones of three sites: Buleji (24° 50' N, 66° 48' E), Hub (24° 54' N, 66° 43' E) and Jiwani (25° 07.630' N, 61° 45.662' E) (Figure 1). Measurements were taken before histological preparations. The animals were brought live to laboratory and the length along their longest axis and shell height of specimens were measured to the nearest millimeter (0.1 mm) by the method described by Galtsoff [13] and Siddiqui and Ahmed [14]. Wet tissue weight (g) was also recorded prior to further examination. Seven (7) to twenty (20) individuals of each species were histologically examined. A portion (5 mm) of soft tissue was excised with a razor blade along a line extending from the lower corner of the labial palps across the stomach to the posterior end of the body. This yielded a transverse section through the stomach, digestive diverticulum, intestine, connective tissue, demibranchs of both gills, and the mantle lobes; tissue samples were then fixed in Davidson's solution [15] for 48 hrs and preserved in 70% ethanol. Prior to sectioning the tissues were dehydrated in an ethanol series, embedded in paraffin wax, sectioned (7µm) and stained with haematoxylin and eosin. Histologically prepared slides of each species were examined for parasitic inclusion following the World Organization for Animal Health guidelines [9, 10]. Gonadal developmental stages were also studies in prepared slides. Gonadal developmental stages

<sup>\*</sup>Address correspondence to this author at the Institute of Marine Science, University of Karachi, Karachi-75270, Pakistan; Tel: +92(21)99261300; E-mail: nuzhatafsar259@hotmail.com

Site	Species	N	Date of Collection
Hub	Crassostrea glomerata	10	8/3/2012
"	Crassostrea madransis	7	24/1/2012 & 14/2/2012
"	Crassostrea gryphoides	20	14/2/2012
Jiwani and Buleji	Saccostrea cucullata	20	2/2/2012 & 21/2/2012
Buleji	Ostrea nomads	7	25/2/2012

Table 1: Sampling Sites, Species, Sample Size and Date of Collection



Figure 1: Map is showing collection sites\*.

were determined by assigning the numeric ranks 0-4 for each gametogenic stage (0=spent, 1=early developing, 2=nearly ripe, 3=ripe, 4=spawning). Scoring scheme was used to assess gonadal condition of different oyster species at different sites to map gonadal variation among species.

# RESULTS

Protozoan *Nematopsis* sp., *Proctoeces* sp. (Trematoda) and some unknown parasites were recorded in *O. nomades* and *S. cucullata* only at Buleji. Calculated biometrics have shown highest shell length (mm) 86.60±7.61 for *C.madransis* followed by 75.60±3.77 *C. gryphoides*, 55.29±0.79 *O. nomads*, 41.60±0.35 *C. glomerata* and collectively 41.05±2.35 for *S. cucullata* obtained from Buleji and Jiwani (Figure

**2**). Meanwhile highest wet weight (g)  $10.6\pm3.8$  was calculated for *C. gryphoides* followed by  $10.2\pm7.6$  *C. madrasensis* and  $5.3\pm2.3$  for *S. cucullata* communally (Figure **2**).

Histological inspection showed specimens of *C. gryphoides* were set up to be almost in resting state. 95.0% individuals were found spent (0) besides one individual was observed with early developmental (1) stage. Whereas *C. madrasensis* was found with 42.9% spent (0) 14.3% early developing (1) and 42.9% in ripe (3) condition. *C. glomerata* showed 10.0% spent (0) 80% nearly ripe (2) and 10% ripe (3) gonads (Figure **2**).

In recent studies individuals of *O. nomades* from Buleji found in ripe (3) state with female gametes and developing (2) male follicles in hermaphroditic individuals. A total of six (6) hermaphroditic and one (1)



Figure 2: Showing average shell length (A); average wet tissue weight (B); gonadal developmental stages of oyster species (C) and parasitic load in different tissues (D). Parasite a-Nematopsis sp.; parasite b-Proctoeces sp. Trematode; parasite c-unknown.

Stages:0=spent; 1=early developing; 2=nearly ripe; 3=ripe;1=spawning.

pure female were encountered during histological investigations. Whereas 15% individuals of *S. cucullata* found developing or nearly ripe (2), 80% ripe (3) and 5.0% spawning (4) observed.

Histological examination has revealed that both of the candidate species were in the verge of the sexually mature state at Buleji and some of these sexually mature individuals have been found with parasitic burden which shows that mature individuals in the wild population have more chances to acquire parasitic burden as during the present study protozoan Nematopsis sp., Proctoeces sp. Trematode parasite and one unknown parasitic cyst were observed in O. nomades and S. cucullata specimens obtained from Buleji. Seven examined slides of O. nomades (one female and six hermaphrodites) were found with protozoan Nematopsis species; among them one hermaphroditic individual was found heavily parasitized with trematode Proctoeces sp. in addition to Nematopsis sp. (Figure 3a, b).

Highest prevalence of infestation were recorded in *O. nomades* at Buleji where *Nematopsis* sp. and *Proctoeces* sp. were found respectively in 100% and 14.8% of individuals, respectively. Furthermore 8% of *S. cucullata* at Buleji were infested with an unknown parasite. Parasitic loads differed between different tissues; *Nematopsis* sp. was found in the gills, digestive and connective tissues in 20.4%, 38.8% and 40.8% respectively of individuals from Buleji. The parasitic load of *Proctoeces* sp. was 44.3% in gills, 14.8% in gonadal portion and 41.0% in connective tissues from the same site (Figure **2**).

A total of thirteen (13) specimens of *S. cucullata* examined for parasitic inclusion from Buleiji and seven (7) individuals from Jiwani were free of any parasitic infection, except for one male specimen that was infected with an unknown parasite, was procured from Buleji. A spherical thraustochytrid-like cyst with honeycomb cellular structures and endospores was observed at the outer periphery of gonadal tissue



b

**Figure 3: a**. Histological sections of *Ostrea nomades*: *A-B* showing sporocysts of *Proctoeces* sp. in the gill filament and the female gonad (mantle), respectively; *C-D* showing Nematopsis sp. in connective tissue and lumen of digestive diverticulum. Infection (black arrows) in gills, gonad, connective tissue and lumen tissues.

**b**. Photomicrograph showing infectious gonadal tissue of *O. nomades* (*E*) and *Saccostrea cucullata* (F-H) male tissue. Arrow points to the *Nematopsis sp.* (*E*) and an unknown (thraustochytrid-like) parasitic cyst (*G*, *H*). Ct=connective tissue; Sc=spermatocytes; Sz=spermatozoa.

(Figure **3b**). In specimens of *C. gryphoides, C. madransis* and *C. glomerata* there was no parasitic inclusion in any specimen examined.

## DISCUSSION

Earlier researchers [14] documented the maximum 241-280 mm size for C. gryphoides at the same site (Hub Delta), the largest size among edible oysters of the world during past decades,. whereas same time the populations of C. gryphoides and C. madrasensis were mostly between 41-200 mm size ranges at Hub Delta. Additionally, larger sizes (40-70 mm) for C. glomerata from Karachi, 32-82 mm for S. cucullata from Paradise Point and 26-97 mm for O. nomads from Buleji and Keamari Sea wall have been recorded formerly [16]. Recent studies have shown the considerable size decrease in existing wild populations when compared to previous data. Although oyster populations at Hub have been considerably depleted during the last decade, any contributing reason behind this is not known so far, but a possible reason which has never been thoroughly investigated in past was the presence of a nuclear power plant at Hub. Nevertheless these species could be useful potential resource for commercial exploitation being acquiring considerable wet tissue mass. Wet tissue weight data for respective species is not available in previous records. However onset of gametogenesis in C. gryphoides in intermediate and large size classes was the same [14]. Further it was indicated that the percentage of females remains lower in the colder months i. e. from November to January and also stated that in winter months more females remain indifferent and undergo resorption. Siddiqui and Ahmed [11] examined the larviparous oyster O. nomads from Buleji and Keamari area and histologically individuals were found to be hermaphroditic, pure male, pure female and indifferent at two sites throughout the year with increasing activity in the winter months followed by immediate ripening and later spawning and resorption around the year.

There is no previous record available pertaining report of trematode parasitic inclusion in Pakistani oyster species. Earlier workers observed 48% to 92% protozoan parasite *Nematopsis* sp. prevalence with an average of 67% in *O. nomads* [5]. Moreover parasitophorous vacuoles containing oocysts were investigated in the digestive gland, palps, mantle, gonads, gills and kidney of the oysters. According to previous report [5] inclusion of *Nematopsis* sp. in *O. nomades* and *C. belcheri* was profound from Keamari Sea Wall constructed near Karachi Port. The presence

of Nematopsis sp. in Buleji population could be due to the fact that Buleji rocky ledge is rich in fauna and flora and number of arthropod species including brachyuran crabs are quite abundant in the area [17]. Oysters are the intermediate hosts of many species of Nematopsis, and life cycle of these parasites is completed in intestine of marine arthropods, moreover species of Xanthidae, Portunidae, Grapsidae, Ocypodidae have been designated by authors which could be the possible final host for pathogenic prevalence [5, 6, 17-19]. Similarly several species of marine brachyuran crabs from Manora Island have been reported [17], which lies adjacent to Keamari sea wall and Buleji. Xanthid crab Leptodious exaratus found to be principal species at both sites. Thalamita prymna, Charybdis species, Eriphia laevimana, Pilumnus vespertilio, Pseudozius caystrus, Grapsus strigosus and Atergatis roseus were also turned out dominant species at both of the sites. These crab species possibly serve as final host for pathogenic infestation.

Identification of observed parasites to species level was not attempted. *Nematopsis* sp. oocysts and metacercaria of the trematode, *Proctoeces* sp. were observed in gills, digestive and connective tissues of *O. nomads* from Buleji. *Nematopsis* has been reported in many bivalve and gastropod mollusks [6, 20, 21]. Boehs *et al.* [6] stated that infestation by sporocysts, which are structures of digenetic trematodes could be a threat for host bivalve because of its potential to reduce host fecundity.

Digenean trematodes have complex life cycles in which bivalve mollusks are intermediate hosts harboring larval stages of the digenean. Some finfish species are known ultimate hosts for bucephalid digenean trematodes [22]. By contrast, Couch [23] observed the un-encysted metacercariae of Proctoeces sp. in the gonadal follicles and ducts of oysters from three Gulf of Mexico estuaries and suggested that the presence of this trematode to its oyster host is in fact not harmful. In the present study metacercariae and mature adults of *Proctoeces* sp. were observed in gills, digestive and connective tissues of gonadal sections in O. nomades. Trematode metacercariae usually cause no apparent harm to host bivalve molluscs [24-26], but infection with Proctoeces sp. may cause serious lesions and inhibit gametogenesis [27].

## CONCLUSION

During the present study protozoan *Nematopsis* sp., trematode *Proctoeces* sp. and an unknown (thraustochytrid-like) parasites have been recorded in

*O. nomades* and *S. cucullata* at Buleji of Sindh coast. On the contrary, Balochistan coast, Pakistan was found to be free of pathogenic infestation during the present oyster watch. There is no previous record of trematode parasites in Pakistani oyster species in literature. In this paucity it is recommended that investigative endeavor ought to be expanded in sustainable deployment perspective.

### REFERENCES

- [1] Mackin JG, Owen HM, Collier A. Preliminary note on the occurrence of a new protistan parasite, Dermocystidium marinum n. sp., in *Crassostrea virginica* (Gmelin). Science III 1950; 328-329. <u>http://dx.doi.org/10.1126/science.111.2883.328</u>
- [2] Alderman DJ. Epizootiology of Mateilia refringens in Europe. Mar Fish Rev 1979; 41(1-2): 67-69.
- [3] Elston RA, Farley CA, Kent ML. Occurrence and significance of bonamiasis in European flat oysters Ostrea edulis in North America. Dis Aquat Org 1986; 2: 49-54. <u>http://dx.doi.org/10.3354/dao002049</u>
- [4] Ford SE, Figueras, AJ. Effects of sublethal infection by the parasite *Haplosporidium nelsoni* (MSX) on gametogenesis, spawning and sex ratios of oysters in Delaware Bay, USA. Dis Aquat Org 1988; 4: 121-133. <u>http://dx.doi.org/10.3354/dao004121</u>
- [5] Siddiqui G, Ayub Z, Sunila I. Prospectus of oyster culture in Pakistan: Pathology assessment of two commercially important oyster species. J Shellfish Res 2008; 27(4): 795-800. <u>http://dx.doi.org/10.2983/0730-</u> 8000(2008)27[795:POOCIP]2.0.CO;2
- [6] Boehs G, Villalba A, Ceuta LI, Luz JO. Parasites of three commercially exploited bivalve mollusk species of the estuarine region of Cachoeira River (Ilheus, Bahia, Brazil). J Invert Pathol 2010; 103(1): 43-47. <u>http://dx.doi.org/10.1016/j.ijp.2009.10.008</u>
- [7] Vico GD, Carella F. Morphological features of the inflammatory response in mollusks. Res Vet Sci 2012; 93(3):1099-1542. <u>http://dx.doi.org/10.1016/j.rvsc.2012.03.014</u>
- [8] Kinne O, Bulnheim HP. International Helgoland Symposium on "Diseases of marine organisms" held on Helgoland from 11th-16th September 1983. Helgoländer Meeresuntersuchungen, 37(1-4). Biologische Anstalt Helgoland: Hamburg 1984; p. 663. ISBN 0017-9957.
- [9] WAOH. Manual of diagnostic tests for aquatic animals. World Organization for Animal Health, Paris 2003; p. 358.
- [10] WAOH. Aquatic animal health code. World Organization for Animal Health, Paris 2007; p. 238.
- [11] Siddiqui G, Ahmed M. Gametogenic patterns of the larviparous oyster *Ostrea nomads* from Karachi, Pakistan (Northern Arabian Sea). Aquacult Res 2002c; 33:1049-1058. http://dx.doi.org/10.1046/j.1365-2109.2002.00769.x
- [12] Siddiqui G. Reproductive pattern of oysters found along the coast of Pakistan (Northern Arabian Sea). World Aquacult Mag 2005; 36: 42-46.

- [13] Galtsoff PS. The American oyster Crassostrea virginica Gmelin. U. S. Fish And Wildlfish Service. Fish Bull 1964; 64(1): 480.
- [14] Siddiqui G, Ahmed M. Histological basis of population structure of three species of *Crassostrea* from Pakistan (Northern Arabian Sea). Pak J Zool 2002b; 34:129-137.
- [15] Shaw BL, Battle HI. The gross and microscopic anatomy of the digestive tract of the oyster *Crassostrea virginica* (Gmelin). Can J Zool 1957; 35(1): 325-347. <u>http://dx.doi.org/10.1139/z57-026</u>
- [16] Siddiqui, G Ahmed M. Oyster species of the sub tropical coast of Pakistan (northern Arabian Sea). Indian J Geo-Mar Sci 2002a; 31(2): 108-118. http://nopr.niscair.res.in/handle/ 123456789/4307
- [17] Siddiqui G, Ahmed M. Distribution and abundance of marine brachyuran crabs on two exposed rocky ledges near Karachi (Pakistan, Arabian Sea). Pak J Zool 1991; 23: 57-63.
- [18] Bower SM, McGladdery SE, Price MI. Synopsis of infectious diseases and parasites of commercially exploited shellfish. Ann Rev Fish Diseas 1994; 4(1): 1-199. <u>http://dx.doi.org/10.1016/0959-8030(94)90028-0</u>
- [19] Ford SE, Tripp MR. Diseases and defense mechanisms. pp. 581-660 in The Eastern Oyster *Crassostrea virginica* (V.S. Kennedy, R.I.E. Newell and A.F. Eble, eds.). College Park: Maryland Sea Grant 1996.
- [20] Lima FC, Abreu MG, Mesquita EFM. Monitoramento histopatologico de mexilhao *Perna perna* da Lagoa de Itaipu, Niteroi. RJ. Arq. Bras. Med. Veterinary Zootech 2001; 53: 203-206.
- [21] Azevedo C, Padovan I. Nematopsis gigas n. sp (Apicomplexa), a parasite of Nerita ascencionis (Gastropoda, Neritidae) from Brazil. J Eukaryot Microbiol 2004; 51(2): 214-219. http://dx.doi.org/10.1111/j.1550-7408.2004.tb00548.x
- [22] Paperna I. Didenea (Phylum Platyhelminthes). In: Woo, P. T. K. (Ed.), Fish Diseases and disorders. Protozoan and Metazoan Infections. University Press, Cambridge 1995; Vol. 1: pp. 329-389.
- [23] Couch JA. Prospective study of infectious and noninfectious diseases in oysters and fishes in three Gulf of Mexico estuaries. Dis Aquat Org 1985; 1: 59-82. <u>http://dx.doi.org/10.3354/dao001059</u>
- [24] Lauckner G. Diseases of Mollusca: Bivalvia. In: Kinne, O. (Ed.), Diseases of Marine Animals. Biologische Anstalt Helgoland Hamburg 1983; pp. 477-879.
- [25] Carballal MJ, Iglesia D, Santamarina J, Ferro-Soto B, Villalba A. Parasites and pathologic conditions of the cockle *Cerastoderma edule* populations of the coast of Galicia (NW Spain). J Invert Pathol 2001; 78(2): 87-97. http://dx.doi.org/10.1006/jipa.2001.5049
- [26] Winstead JT, Volety AK, Tolley SG. Parasitic and symbiotic fauna in oysters (*Crassostrea virginica*) collected from the Caloosahatchee River and estuary in Florida. J Shellfish Res 2004; 23(3): 831-840.
- [27] Sunila I, Williams L, Russo S, Getchis T. Reproduction and pathology of blue mussels, *Mytilus edulis* (L.) in an experimental longline in Long Island Sound, Connecticut. J Shellfish Res 2004; 23: 731-740.

Received on 31-03-2014

Accepted on 21-05-2014

Published on 03-06-2014

© 2014 Afsar et al.; Licensee Lifescience Global.

http://dx.doi.org/10.6000/1927-5129.2014.10.30

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<u>http://creativecommons.org/licenses/by-nc/3.0/</u>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.