

Toxic Effects Observed on Light Weight Proteins of *Musca domestica* with Pb(CH₃COO)₂

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Abstract: Lead is considered to be an important noxious waste which could contaminate the environment, such as soil, air and water, therefore, insects could be influenced easily by the lead. *Musca domestica*, was studied at 48 hours post treatment, under the effects of lead acetate, in different concentrations. It was observed that under the effects of lead abnormalities and deformity were developed in the larvae of flies. Thus these flies could present a useful module for the quick transmission of the environmental hazards due to lead contamination, which exerts a specific physiological and morphological effect on these flies.

Keywords: Effects, lead acetate, *M.domestica*.

INTRODUCTION

Lead is an important environmental pollutant, due to the possible hazardous effects of this metal, the detection of its presence in the environment is therefore a matter of urgent concern. Although, many studies have been carried out in relation with the biological effects of lead, its toxic potential against insects remains to be established. Though [1-8b, 9, 10] have reported cytological effects of this metal leading to expected morphological abnormalities. Some studies have been carried out on natural populations of *Musca domestica* in respect of effects of heavy metals, it has been established that contamination with heavy metals (Zinc, Lead etc.) can induce the effects on feeding behavior of flies. Their morphological abnormalities, functional modifications and malformations could be induced under heavy metal actions [11, 12]. [8a] indicated that induced abnormalities could be obtained due to the effect on meiotic nondisjunction Rizwan *et al.* [13, 14] have reported morphological effects of this metal on insects and suggested to use these insects as indicators of the presence of this metal in the environment. It lives feeding on garbage, specially where sanitary conditions are unsatisfactory [15-17]. Since it is found all over the world, therefore, it could act as a model for the detection system of pollution with this heavy metal.

Among the heavy metals, lead, has been shown to be widely distributed in the atmosphere, water, soils

and foods [18]. lead inhibits the activity of enzymes that are dependant on the presence of free sulphhydryl groups (SH). The clearest manifestation of these effects is the disturbance on the biosynthesis of heme, which in humans is accompanied by abnormalities in porphyrin metabolism [19]. Lead acetate is used as a topical astringent and is found to be a renal carcinogen in rats [20-25]. In the Syrian hamster lead induces neoplastic changes in the bronchio-alveolar area [26, 27]. It also produces infertility in mice [28] and reduces the reproductive ability of rats [29-31]. In *Drosophila melanogaster* lead induces enzymatic alterations in esterase and triose phosphate isomerase [32] and affects non disjunction [8a]. However, information about the mutagenic effects of lead salts in humans who are occupationally exposed to them and information obtained from *in vitro* studies are contradictor [33].

Electrophoresis is being broadly used for categorization of proteins and peptides for the diagnostic and/ or preparative unification of organic macromolecules [34]. The process of electrophoresis first used by [35] for the separation of proteins has found many dimensions in analyzing and separating macromolecules. These techniques, wheather alone or in combination, have proved to be very useful for proteins and peptides and the complex proteome analysis [36].

MATERIALS AND METHODS

For obtaining an initial wild house flies culture, a usual larval media for *Musca domestica*, was prepared

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Solution	Preparation
i) Acrylamide-Bisacrylamide solution(30.0:0.8)	Dissolve 30 gm acrylamide and 0.8 gm bisacrylamide in deionized water. Make up the volume to 100 ml. Filter through Whatman no. 1.
ii) 1.5 M Tris-HCl buffer:	Tris 18.02 gm, dissolve in 80 ml and adjust the pH of this solution to 8.8 using 0.1M HCl. Make up the volume to 100 ml with deionized water.
iii) 10% Sodium dodecyl sulfate:	Dissolve 1 gm SDS in 9 ml water and make the volume up to 10 ml with deionized water.
iv) 10% Ammonium persulfate:	Dissolve 1 gm APS in 1 ml water and make the volume up to 10 ml with deionized water
v) Sample diluting buffer (SDB):	Dissolve 6.25 ml of 1M Tris-HCl pH 6.8 (Solution C), 2 gm SDS, 5 ml 2-mercaptoethanol and 10 ml glycerol together. Make volume up to 100 ml with deionized water.
vi) Reservoir Buffer:	Dissolve 0.9 gm Tris, 3.6 gm Glycine and 1.0 gm SDS in 500 ml deionized water. Make up to 1 liter.
vii) Staining solution: (Bromophenol Blue and 0.2% Coomassie blue).	Dissolve 0.5 gm Coomassie blue in 18.75 ml acetic acid and 12 ml methanol, Make volume up to 2.50 ml with
viii) Destaining solution	Mix 10 ml Acetic acid and 30 ml methanol. Make up the volume to 100 ml.

Figure 1: Preparation of solutions.

in a wide plain pot by mixing wheat bran with milk and sugar in a ratio of 10: 3: 3 respectively, some amount of water added to make it a uniform mixture in a texture of horse dung and put it in an open place for wondering.

House flies freely to visit thereon. Thereafter, the pots were brought into the laboratory and kept for 2 to 3 days in a cage for egg hatching. Housefly larvae were reared under aseptic conditions on a basic diet with amended procedure described by [37]. Larvae took around 6 days to become full grown, full grown larvae were removed from the medium and allowed to pupate in separate covered glass bottles at 29-30 °C. After having established a good supply of desired aged insects, early 3rd instar larvae were drawn out and treated in the batches of 10 each in separate culture bottles, supplies with 3 grams bananas mixed with desired amount of lead acetate. Doses of lead acetate were used as, 0.25 mg, 0.5 mg, and 1.0 mg. A batch of untreated three bottles was kept as control. larvae were exposed in each bottle for 48 hours. After that mortality of larvae in each bottle was observed. Surviving larvae were kept in separate bottles on lead free banana diet up to emergence. During that period effects of lead acetate on pupation and adults of under test insect was observed.

The determination of lead acetate on protein of *Bactrocera cucurbitae* larvae were studied with lead acetate kept for 48 hours exposure. Thereafter, crushing and homogenizing of the treated and untreated larvae was made. Preparation of solution,

Reagent and chemicals are shown in Figures 1 and 2 respectively.

Reagents and Chemicals	Brand
Acrylamide	Fluka
N,N,Methylene bisacrylamide	Fluka
Tris (hydroxymethyl) aminomethane	Fluka
HCL	Merck
Sodium dodecylsulfate	Fluka
Ammonium persulfate	Merck
Glycine	Fluka
TEMED	Merck
Bromophenol blue	Merck

Figure 2: Reagents and Chemicals.

Preparation of Gel

In the process of electrophoresis, the capillary tubes of electrophoresis were cleaned by water and ethanol then dried it by air. The lower mouth of capillaries were covered by rubber stopper. 10 ml resolving gel was prepared with above ingredient. The mix solution was filled in capillaries tube, then added the 0.1 ml ammonium sulphate and 0.008 ml TEMED in capillaries, then left it for 3-4 hours for polymerization, after that 200µl. (micro litre) sample was added and then Bromophenol solution was added. After 30-40 min. the mouth of above and lower part of capillaries were exposed with Reservoir Buffer solution in the electrophoresis tank for one day under 110 volt. After that gel were exposed to coomassie blue solution for 2 hours, after colorization of Gel, It was kept in the

Table 1: Values of Various Proteins Observed in Lead Acetate Treated and Untreated *Musca domestica* Larvae

Rf	Egg Albumin control	<i>Musca domestica</i> Normal (untreated)	<i>Musca domestica</i> treated
0.04	+	-	-
0.71		-	+
0.72		+	-
0.81		+	-
0.85		-	+
0.93		+	-
0.96		-	+

Protein: + = present - = absent Rf= Relative flow.

destaining solution for removing the excess color on the Gel then the bands of proteins were observed. After this process the length and bands on Gel was measured for Rf determination. Egg albumin was also run simultaneously, for the comparison.

RESULTS

The relative flow (rf) of the protein of *Musca domestica* (untreated) were found to be 0.46, 0.53, 0.72, 0.81, 0.93. On the other hand the relative flow (rf) of Protein of *Musca domestica* (treated) indicated the values as 0.53, 0.61, 0.71, 0.85, and 0.96, respectively.

As shown in (Table 1) Protein rf. 0.72, 0.81, and 0.93, have not been observed in treated *Musca domestica*. While protein relative flow (rf) 0.71, 0.85 and 0.96, have been detected as altered in *Musca domestica*.

Electrophoretic expression of various proteins flow as compared to egg albumin in treated and untreated *Musca domestica* shown in (Figure 3).

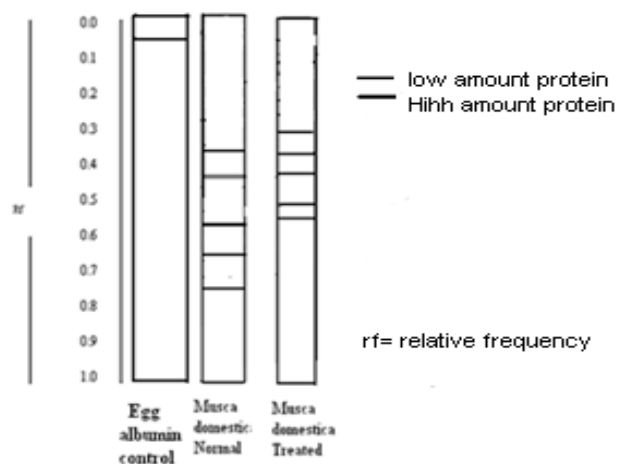


Figure 3: Electrophoretic expression of various proteins flow as compared to egg albumin in treated and untreated *Musca domestica*.

DISCUSSION

Protein relative flow (rf 0.71) was found in *Musca domestica* (treated) that is seems to be lighter than the egg albumin, while corresponding protein, in the untreated *Musca domestica*, was absent at the same rf. This suggests that the protein (rf 0.61) was affected with some alteration in insect. Protein (rf 0.72) was found in *Musca domestica* (untreated) that is seems to be lighter than the egg albumin, while corresponding protein, in the treated *Musca domestica*, was absent at the same rf. This suggests that protein (rf 0.72) was affected with some alteration in the untreated insect. In the *Musca domestica* protein (rf 0.81) was found in the untreated ones while it is absent in the treated. That suggest the protein (rf 0.81) was affected at a low extend. Protein LII (rf 0.85) was found in *Musca domestica* (treated) that is seems to be lighter than the egg albumin, while corresponding protein, in the untreated *Musca domestica*, was absent at the same rf. This suggests the protein (rf 0.85) was affected with some extent in the treated insect. Protein (rf 0.93) was found in *Musca domestica* (untreated) that is seems to be lighter than the egg albumin, while corresponding protein, in the treated ones, were absent at the same rf. This suggests that the protein (rf 0.93) was affected with some extent in the untreated insect. Protein (rf 0.96) was found in *Musca domestica* (treated) that is seems to be lighter than the egg albumin, while corresponding protein, in the untreated *Musca domestica*, was absent at the same rf. This suggests that the protein (rf 0.96) was affected with some extent in the treated insect.

Drosophila melanogaster treated with different doses of lead acetate viz. 0.125 mg., 0.25 mg., 0.5 mg., 1.0 mg and 2.0 mg resulted deformities. [38], indicated cellular damage in processes of lead exposed to PC-12 cells. [39], reported that heavy metal compounds divergence is evidently a extensive

observable fact in invertebrates. After lead exposure the N-acetylcysteine (NAC), glutathione (GSH), glutathione disulfide (GSSG) and malondialdehyde (MDA), were found effected after treated to various doses of lead acetate, these results could be correlated with the present findings with the presence of affected proteins in the lead treated insects. [40], indicated that, lead is a pollutant heavy metal, which can be absorbed by the digestive system in a 10%, [41], indicated that when lead incorporated by cells, it produces free radicals, H_2O_2 and $\cdot OH$. [42], found free radicals can also produce simple breaks in the DNA chains these results resembled with present finding that exposure of lead produced the abnormal morphological effects in the larvae and the adults emerged therefrom [43] reported newly hatched nymphs of an Indian short horned grasshopper *Oxya fuscovittata* (Marschall) Orthoptera: Acrididae were fed on foods treated with three sub lethal concentrations of CdCl i.e. 2. 25 ppm in oat or dose 1 (d1), 50 ppm in oat or dose2 (d2) and 100 ppm in oat or dose3 (d3) until they reached on adult stage for a complete generation. Growth was measured in terms of specific growth rate (SGR), average daily growth (ADG), percent weight gain (PWG) and Growth rate (GR). They observed that growth retardation occurred significantly with the increase of doses in both sexes. Adult life period found reduced in both sexes however, in females a significant difference was found only with higher doses (d2 and d3). Lower survival d3 was observed. These adverse effects of heavy metals on diptera are in the line with the present findings. [44] found morphological changes in wild *Drosophila* species that found over almost all of Europe, under the effects of lead. The effects of lead on inversion polymorphism were studied by cytological analysis of gene arrangements on all of the five acrocentric chromosomes, as well as by cytological analysis of karyotypes on all of the four autosomes. The frequencies of particular gene arrangements on the four autosomes changed significantly in the samples maintained on medium not supplemented with lead. The frequencies of some gene arrangements on all of the five acrocentric chromosomes changed significantly in the flies maintained on media supplemented with lead. The length of exposure to different lead concentrations results in a significant change in the frequency of a few gene arrangements on two autosomes. Their results showed that different concentrations of lead, and exposure period caused affects on chromosome the effects on the DNA configuration and chromosome cause effects on morphology and the physiology of the affected

organism, in this way presently the obtaining of altered protein bands, deform larvae, pupae and deform adults are in the line with the previous findings.

The present observation showed that lead acetate induced there effects mostly on the morphology and development of the under test insects. On the other hand, the differences due to increase in concentration of lead acetate influenced various changes as compared to control.

REFERENCES

- [1] Tachi K, Nishime S. Cytogenetic effects of lead acetate on rat bone marrow cells. *Arch Environ Heal* 1975; 403: 144-47.
- [2] Michailova P. Comparative karyological studies of three species of the genus *Glyptotendipis* Kieff. (Diptera, Chironomidae) from Hungary and Bulgaria and *Glyptotendipis salinus* sp. n. from Bulgaria. *Folia Biol (Krakwo)* 1987b; 35: 43-56.
- [3] Short C. Varion in sister-chromatid exchange among 100 species of the general insect population. *Proc Environ* 1990; 29: 140-149.
- [4] Wilson BS. Sister chromatid exchange in larvae of insects. *Lab* 1995; 62: 135-44.
- [5] Watson N. Chromosome aberrations and sister chromatid exchanges in insects of lead polluted. *Exp Sci* 1999; 90: 64-69.
- [6] Walter R. Cyntheses prozesse an den Riesenchromosomen Von. *Glyptotendipes J Sci* 2000; 128: 80-85.
- [7] Porter DM. Mutagenicity new horizons in genetic toxicology. *Proc Tox* 2002; 55: 32-37.
- [8] (a) Ramel C. Effects of metal compounds on chromosome aggregation. *Mutat Res* 1973; 21: 45-46; (b) Ramel S. Chromosomal aberrations in insects. *Environ Sci* 2003; 16: 119-27.
[http://dx.doi.org/10.1016/0165-7992\(73\)90062-6](http://dx.doi.org/10.1016/0165-7992(73)90062-6)
- [9] Talbot PS. Sister-chromatid exchange frequency correlated with age, sex and lead poisoning. *Environ Tox* 2004; 25: 27-33.
- [10] Margim A. Chromosome affected in experimental lead poisoning. *Tox* 2005; 41: 6-14.
- [11] Michailova MP. The effect of metal compounds on chromosome segregation. *Ist Nat Conf Plovidiv* 1987a; pp.168-173.
- [12] Timmermans KP. Heavy metal body burden in insects larvae as related to their feeding behaviour. *Symp Abst* 1988; 76.
- [13] Rizwan-ul-Haq, Farhanullah KM, Ehtesham-ul Haq. Teratogenic effect of lead acetate on *Bactrocera cucurbitae*(COQ.). *Pak Entomol* 2011 (b); 33(1): 41-4.
- [14] Rizwan-ul-Haq, Farhanullah KM, Ehtesham-ul Haq. Adverse effect of lead acetate on *Drosophila melanogaster*. *J Basic Appl Sci* 2011(a); 7(2): 157-63.
- [15] Greenberg B. Flies and diseases. *Biology and Disease Transmission*. Princeton Univ. Press, Princeton. 1973; Vol. 3.
- [16] Graczyk TK, Knight R, Gilman RH, Cranfield MR. The role of non-biting flies in the epidemiology of human infectious diseases. *Microb Infect* 2001; 3: 231-35.
[http://dx.doi.org/10.1016/S1286-4579\(01\)01371-5](http://dx.doi.org/10.1016/S1286-4579(01)01371-5)
- [17] Graczyk TK, Knight R, Tamang L. Mechanical transmission of human protozoan parasites by insects. *Clin Microbiol Rev* 2005; 128-132.
<http://dx.doi.org/10.1128/CMR.18.1.128-132.2005>

- [18] Beliles RP. Metals. In *Toxicology*. Casarett LJ, Doull J, Eds. Macmillan Pub. New York 1975; pp. 477-482.
- [19] Valle BL, Ulmer DD. Biochemical effects of mercury, cadmium and lead. *Ann Rev Biochem* 1972; 41: 91-128. <http://dx.doi.org/10.1146/annurev.bi.41.070172.000515>
- [20] Boyland E, Dukes CE, Grover PL, Mitchley BCV. The induction of renal tumor by feeding lead acetate to rats. *Br J Cancer* 1962; 16: 283-88. <http://dx.doi.org/10.1038/bjc.1962.33>
- [21] Van Esch GJ, Gendersen H, Vink HH. The induction of renal tumors by feeding of basic lead acetate to rats. *Br J Cancer* 1962; 16: 289-97. <http://dx.doi.org/10.1038/bjc.1962.34>
- [22] Roe FJC, Boyland E, Dukes CE, Mitchley BCV. Failure of testosterone or xanthopterin to influence the induction of renal neoplasms by lead in rats. *Br J Cancer* 1965; 19: 860-66. <http://dx.doi.org/10.1038/bjc.1965.99>
- [23] Mao P, Molnar JJ. The fine structure and histochemistry of lead induced renal tumors in rats. *Am J Pathol* 1967; 50: 571-80.
- [24] Choie DD, Richter GW. Cell proliferation in mouse kidney induced by lead. 1 Synthesis of DNA. *Lab Invest* 1974; 30: 647-51.
- [25] Furst A, Schnauzer M, Sasmore DP. Tumorigenic activity of lead chromate. *Cancer Res* 1976; 36: 1779-83.
- [26] Kobayashi N, Okamoto. Effects of lead oxides on the induction of lung tumours in Syrian Hamster. *J Natl Cancer Inst* 1974; 52: 1605-607.
- [27] ICPEMC. Report of ICPEMC task group 5 on the differentiation between genotoxic and non-genotoxic carcinogens. *Mutat Res* 1984; 133: 1-49.
- [28] Varma MM, Joshi SR, Adeyami AO. Mutagenicity and infertility following administration to lead sub-acetate to Swiss male mice. *Experientia* 1974; 30: 486-87. <http://dx.doi.org/10.1007/BF01926307>
- [29] Stowe HD, Goyer RA. The reproductive ability and progeny of F₁ lead toxic rats. *Fertility Sterility* 1971; 22P 755-60.
- [30] Hackett PL, Hess JO, Sikov MR. Effect of dose level and pregnancy on the distribution and toxicity of intravenous lead in rats. *J Toxicol Environ Health* 1983; 9: 1007-20. <http://dx.doi.org/10.1080/15287398209530221>
- [31] Hess JO, Sikov MR. Distribution and effects of intravenous lead in the fetoplacental unit of the rat. *J Toxicol Environ Health* 1982; 9: 1021-32. <http://dx.doi.org/10.1080/15287398209530222>
- [32] Lower WF, Drobney VK, Rose PS, Putnam CW. Environmental and laboratory monitoring of biotic indicators of heavy metals. *Mutat Res* 1976; 38: 386.
- [33] Maki-Paakkanen JM, Sorsa M, Vainio H. Chromosome aberrations and sister chromatid exchanges in lead exposed workers. *Hereditas* 1981; 94: 269-75. <http://dx.doi.org/10.1111/j.1601-5223.1981.tb01764.x>
- [34] Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T₄. *Nature* 1970; 227: 680. <http://dx.doi.org/10.1038/227680a0>
- [35] Tiselius A. A new apparatus for electrophoretic analysis of colloidal mixtures. *Trans Faraday Soc* 1937; 33: 524. <http://dx.doi.org/10.1039/tf9373300524>
- [36] Andrew AT. *Electrophoresis: Theory, techniques and biochemical and clinical applications*. Clarendon Press, Oxford 1986.
- [37] Naqvi SN H, Tabassum M, Jahan N, Yasmeen MA, Azmi Matin Z. Toxicity and abnormalities produced by instar larvae of *Musca domestica* L. *Pro Pakistan Congr Zool* 1994; 14: 283-290.
- [38] Nukhet AB, Elizabeth A, Franklin NE. Effects of N-acetylcysteine on lead-exposed PC-12 cells. *Arch Environ Contam Toxicol USA* 2005; 49: 119-23. <http://dx.doi.org/10.1007/s00244-004-0025-0>
- [39] Ahmed SO, Naqvi SNH. Toxicity and effects of Dimilin on protein pattern of *Aedes aegypti*. *Proc Entomol Soc* 1985; 14&15: 119-32.
- [40] Corey OG, Galvao CL, Plomo. Serie Vigilancia 8. Metepec, Edo. De México. Centro Panamericano de Ecología Humanay Salud Org Panam Salud O M S 1989; p. 103.
- [41] Roy NK. Mutagenesis and comutagenesis by lead compounds. *Res* 1992; 298: 97-103.
- [42] Friedberg EC, Walker G, Siede W. *DNA Repair and Mutagenesis*. ASM Press, Washington, 1995; pp. 16-17.
- [43] Chandrik M, Arijit G, Parimalendu H. Influence of Cadmium on Growth, Survival and Clutch Size of A Common Indian Short Horned Grasshopper, *Oxya fuscovittata*. *Am-Eurasian J Toxicol Sci* 2009; 1(1): 32-36.
- [44] Kalajdzic P, Stamenkovic-Radak M, Andjelkovic M. The effect of different concentrations of lead on inversion polymorphism in *Drosophila subobscura*. *Hereditas* 2006; 143: 41-6. <http://dx.doi.org/10.1111/j.2006.0018-0661.01939.x>

Received on 16-06-2012

Accepted on 02-07-2012

Published on 20-07-2012

<http://dx.doi.org/10.6000/1927-5129.2012.08.02.21>

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